



# In vitro monitoring of natural thorium in urine using fluorimeter

S. Kalaiselvan<sup>a,\*</sup>, A.R. Lakshmanan<sup>b</sup>, V. Meenakshisundaram<sup>a</sup>

<sup>a</sup> Radiological Safety Division, Indira Gandhi Centre for Atomic Research, Kalpakkam 603 102, India

<sup>b</sup> Savitha Engineering College, Thandalam, Chennai 602 105, India

## ARTICLE INFO

### Article history:

Received 18 June 2011

Received in revised form

15 September 2011

Accepted 16 September 2011

Available online 1 October 2011

### Keywords:

Bioassay

Thorium

Solvent extraction

Fluorimetry

## ABSTRACT

A relatively less expensive and less time consuming radio analytical technique for quantitative determination of  $\text{Th}_{\text{nat}}$  in urine at mBq level is developed and reported in this paper. Th in urine is co-precipitated with  $\text{Ca}_3(\text{PO}_4)_2$  from wet oxidized urine matrix and the precipitate is dissolved in  $\text{HNO}_3$  and evaporated to dryness. The residue is dissolved in 3 M HCl and 200 mg of Na-EDTA is added to mask  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$  and  $\text{Fe}^{3+}$  ions.  $\text{Th}^{4+}$  is extracted into 0.01 M PC-88A (2-ethyl hexyl phosphonic acid mono-2-ethylhexyl ester), dissolved in toluene from the experimentally optimized pH  $2.5 \pm 0.3$  in aqueous phase.  $\text{Th}^{4+}$  is stripped into 8.0 M HCl and evaporated to dryness. The content of the beaker is dissolved in pH 1.8 HCl and complexed with 3-hydroxy flavone. The sample is excited at 397 nm and fluorescence intensity is measured at 462 nm. The detailed study of the method is presented in this paper. Interference study on elements that are normally present in urine and other actinides (if present) is also given.

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## 1. Introduction

Radiation workers who handle thorium are routinely monitored at predetermined intervals. After the intake, the majority of thorium that enters the body is excreted through the feces in the first 48 h and a small amount through the urine. Urine containing thorium undergoes systemic excretion whereas feces containing thorium undergoes both systemic and non-systemic excretions. Therefore, urinalysis is preferred for dose assessment. In addition, the body also has a retention factor for thorium which drops rapidly [1]. Hence, sensitive and fast method is required to estimate thorium in urine. Sensitive techniques like ICPMS method with MDA 1.0 ng/l [2] and Neutron Activation Analysis (NAA) with MDA 0.01 ng/l [3] are very much suitable for low level estimation of natural thorium but are expensive. It is reported that alpha spectrometric technique involving anion exchange separation requires seven days for completion of analysis with MDA 0.1 mBq/l [4]. Hence, an attempt is made to develop a sensitive and fast method for estimation of intake of thorium and the same is given in detail in this paper. This is a solvent extraction method of using PC-88A as extractant [5–7] and estimation of quantity using fluorimeter. Thorium is extracted into 2-ethyl hexyl phosphonic acid mono-2-ethyl hexyl ester (PC-88A) dissolved in toluene. In addition, fluorescent technique is also optimized to determine thorium at sub microgram

level by complexing with 3-hydroxy flavone dissolved in different polar solvents.

## 2. Experimental

### 2.1. Chemicals

All chemicals used in the experiments including  $\text{Th}(\text{NO}_3)_4 \cdot 4\text{H}_2\text{O}$  were of analytical reagent grade.

### 2.2. Instrument

Fluorolog-3 Jobin Yvon–Spex Spectrofluorimeter, New Jersey is used. Its spectral resolution is 0.2 nm.

### 2.3. Method

The separation scheme for thorium from urine matrix is given with flow chart format in Fig. 1. The scheme essentially consists of three steps viz: (a) pre concentration, (b) solvent extraction separation and (c) fluorimetric estimation.

#### 2.3.1. Pre-concentration

To 1000 ml of each pooled urine sample, a known amount of Th at microgram level was spiked, stirred and heated with 50 ml of conc.  $\text{HNO}_3$ , 25 ml of conc. HCl and 1 ml of  $\text{H}_2\text{O}_2$  until the solution turns into pale yellow. Thorium is then co-precipitated with  $\text{Ca}_3(\text{PO}_4)_2$  by adjusting the pH of the sample to 9 using ammonia. The precipitate is centrifuged and dissolved in 10 ml of 8 M

\* Corresponding author. Tel.: +91 044 27480062; fax: +91 044 27480235.  
E-mail address: [kalai@igcar.gov.in](mailto:kalai@igcar.gov.in) (S. Kalaiselvan).

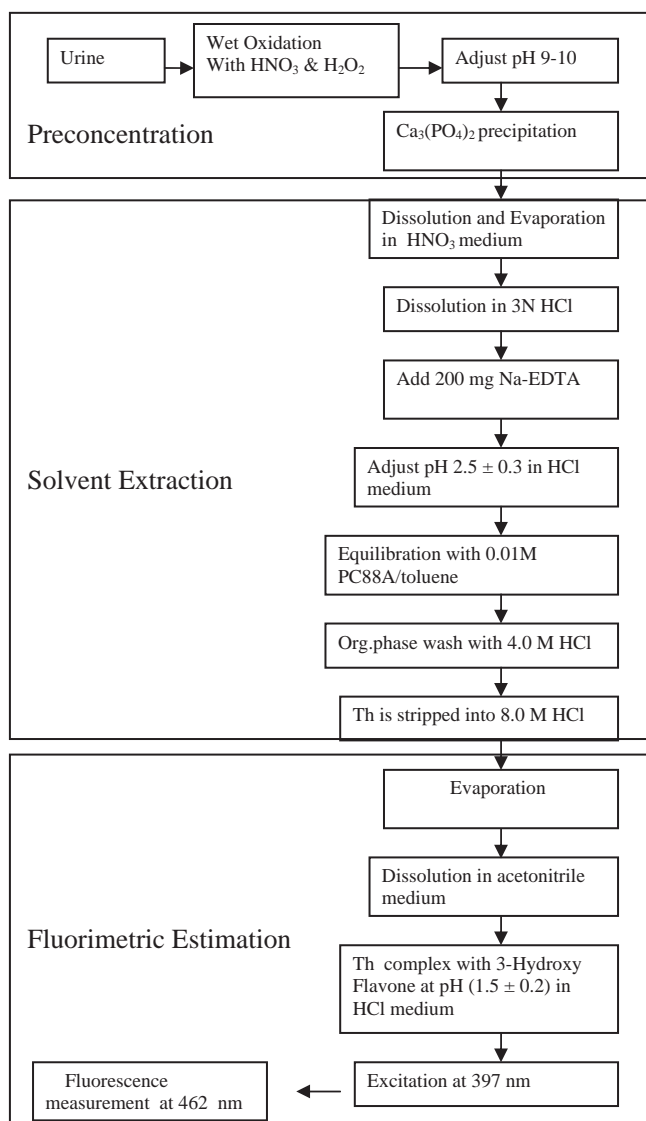


Fig. 1. Flow chart of separation schemes for Th in urine matrix.

HNO<sub>3</sub> followed by evaporation to destruct organic matter. Finally, the residue is dissolved in 10 ml of 3 M HCl and the solution is covered with a watch glass and boiled for 30 min to hydrolyse the pyrophosphate and to get homogeneous solution.

### 2.3.2. Solvent extraction separation

To the pre-concentrated solution, 200 mg Na-EDTA is added and the pH is adjusted to  $2.5 \pm 0.3$  using 10% ammonia and 6 M HCl. The sample solution is filtered and the filtrate is equilibrated with 15 ml of 0.01 M PC-88A/toluene for 10 min. This step is repeated with fresh PC-88A for quantitative extraction of Th. Then the organic phase is collectively washed with 4 M HCl to remove U [7]. Finally, the organic phase is equilibrated with 15 ml of 8.0 M HCl for 10 min. This step is repeated with fresh 8 M HCl solution for quantitative elution of Th into HCl solution [7] and evaporated to dryness.

### 2.3.3. Fluorimetric estimation

The residue content in the beaker is dissolved in 1.5 ml volume of HCl solution (pH 1.8), transferred to 10 ml standard flask, added 1 ml of 0.01% 3-hydroxy flavone (prepared in acetonitrile solvent) and the rest of the volume is made up with acetonitrile solvent. The pH of the resultant solution is  $1.5 \pm 0.2$ . The solution is taken in the cell and kept in the sample holder of the fluorimeter. The

sample excited at 397 nm and fluorescence intensity is measured at 462 nm.

## 3. Results and discussion

### 3.1. Optimization of the conditions for fluorimetric estimation of Th

The complex of thorium with morin, quercetin and 3-hydroxy flavone (3-HF) gives fluorescence when they are excited at 420 nm, 425 nm and 390 nm respectively. The Minimum Detectable Activity (MDA) for Th when complexes with Morin and Quercetin is 20 µg/l [8,9] and for 3-hydroxy flavone, it is 10 µg/l [10]. It is also given in the literature that the fluorescence of flavone complexes with metals are higher in polar solvents than in apolar solvents. Therefore, to arrive at maximum fluorescence intensity for a given amount of Th, the complex of thorium-3-hydroxy flavone was investigated using different polar solvents like ethanol, methanol, acetonitrile, dioxane, isopropanol and in tetrahydrofuran.

The stability of the complex depends essentially on pH. Hence, optimization studies on the pH value for thorium-3-hydroxy flavone complex for given solvents were undertaken. The pH value of thorium-3-hydroxy flavone complex was varied between 0 and 3.5 in different polar solvents and the fluorescence intensity of the complex was measured for a known amount of Th (i.e. 9.45 µg/ml). The complex was excited at 397 nm and the emission peak was observed between 461 and 467 nm for different polar solvents as shown in Table 1. It is found that among various solvents, the ratio of the average fluorescence intensity due to Th with respect to average fluorescence intensity due to blank is the highest (i.e. 892) in acetonitrile solvent. The relative fluorescence intensity of the complex in different polar solvents is given in Fig. 2. The optimum pH value of the complex for maximum fluorescence efficiency was observed between 1.5 and 1.8. Among different polar solvents, maximum fluorescence intensity was observed in methanol and ethanol solvents when the concentration of Th is 9.45 µg/ml and no fluorescence is observed in both the solvents when Th concentration is decreased to 0.5 µg/ml. However, in acetonitrile solvent, fluorescence is observed at this concentration of Th.

### 3.2. Calibration of fluorimeter

To estimate the percent chemical recovery of Th from the above established method, fluorimeter needs to be calibrated. To calibrate fluorimeter, fluorescence intensity of Th-3-hydroxy flavone was measured for different amounts of Th<sub>nat</sub> ranging from 0.4 to 4.8 µg spiked in each 10 ml volume. Measurements were taken in both right-angle and front-face modes. From the data obtained as given in Table 2, it was observed that the best fitted line could be obtained in the right angle mode with slit width 2.0 nm both for excitation and emission monochromator (Fig. 3). A typical fluorescence spectrum obtained in comparison with blank (zero amount of Th) is shown in Fig. 4. From this, it can be inferred that low level of thorium can quantitatively be estimated which is the objective of developing this method.

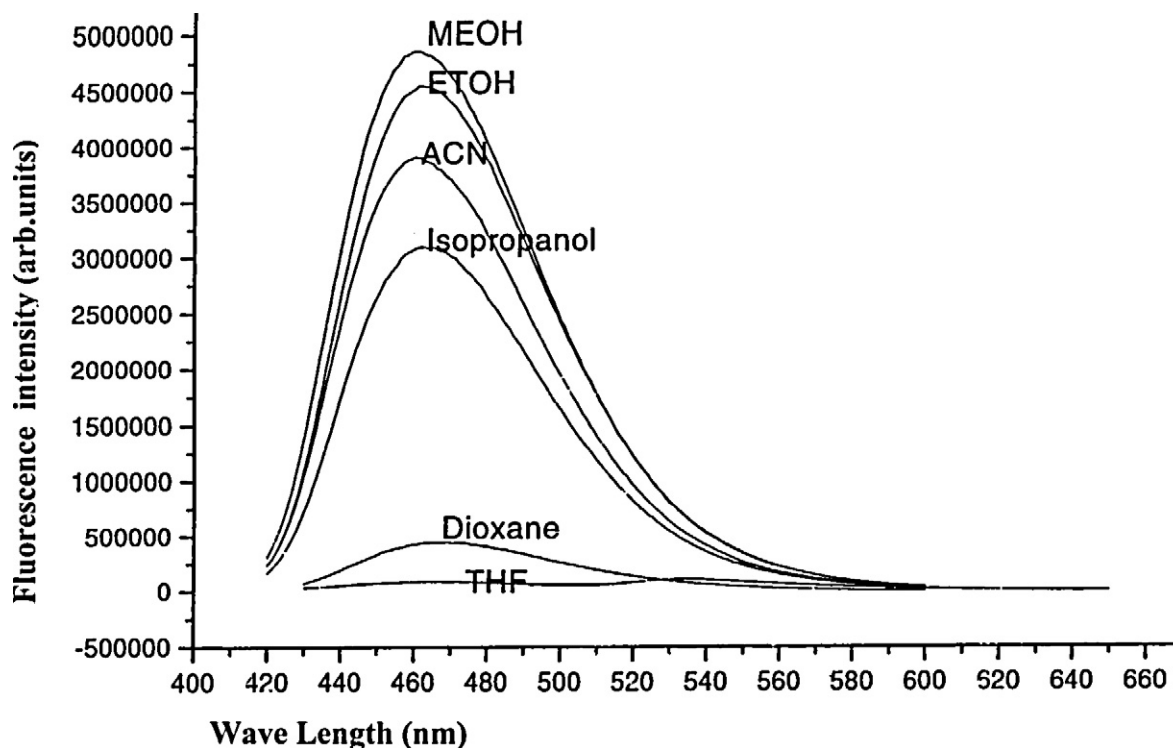
### 3.3. Optimizing the conditions for extraction of Th into PC-88A

It is reported in the literature that Th can be quantitatively extracted from dilute HClO<sub>4</sub> solution in the pH range 2.3–3.8 with 0.01 M PC-88A dissolved in toluene [7]. But the presence of Ca<sup>2+</sup> and Mg<sup>2+</sup> ions each at 100 mg and PO<sub>4</sub><sup>3-</sup> ions at 500 mg in the aqueous phase gave undesirable precipitate during equilibration with PC-88A. Therefore, experiments were repeated with different acidity of aqueous phase using dil. HCl from pH 0 to 5.0 and found that Th could quantitatively be extracted between pH 2.5 and 3.5.

**Table 1**

Fluorescent intensity of Thorium-3-hydroxy flavone complex with different polar solvents at microgram level of Natural Thorium (9.45 µg/ml).

Name of solvent	Optimized pH of the complex	Excitation wavelength (nm)	Emission wavelength (nm)	Average fluorescent intensity due to Th (A)	Average fluorescent intensity due to blank (B)	Th/Blank A/B
Tetra hydrofuran	1.5 ± 0.2	397	467	$8.7 \times 10^3$	9385	0.92
Isopropanol	1.8 ± 0.2	397	463	$31 \times 10^5$	879 510	3.52
Ethanol	1.8 ± 0.2	397	461	$46 \times 10^5$	19 907	231.11
Dioxane	1.5 ± 0.2	397	467	$45 \times 10^4$	12 345	36.45
Methanol	1.6 ± 0.2	397	461	$49 \times 10^5$	277 734	17.64
Acetonitrile	1.5 ± 0.2	397	462	$39 \times 10^5$	4373	891.84

**Fig. 2.** Relative fluorescence intensity of Th-3-hydroxy flavone at 9.45 µg/ml of Natural Thorium.

The amount of  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  ions present in 24-h urine sample varies in the range 100–300 mg, 100–200 mg [11]. It was found that up to 300 mg  $\text{Ca}^{2+}$ , and 300 mg  $\text{Mg}^{2+}$  ions present together in the aqueous phase of dil. HCl in the pH range 2.5–3.5 did not interfere in the extraction of Th with PC-88A. However, the presence of  $\text{Fe}^{3+}$  ions even at 1.0 mg level gets precipitated as  $\text{FePO}_4$ . It was found that the presence of Fe even at 10 mg level was effectively masked by 200 mg Na-EDTA.

### 3.4. Validation of the method

Urine samples collected from unexposed personnel were divided into 12 equal parts of each 1 l and in each sample, a known amount of  $\text{Th}_{\text{nat}}$  (4.0 µg) was spiked. Each set consists of six samples. One set was processed by standard anion exchange method with spectrophotometric estimation [12] and the other set by the present developed method. Average recovery obtained by anion

**Table 2**

Linearity of Th-3-hydroxy flavone complex in acetonitrile solvent.

Amount of Th (µg/10 ml)	Right angle mode		Front face mode	
	Slit width 1.0 nm	Slit width 2.0 nm	Slit width 1.0 nm	Slit width 2.0 nm
0	0	0	0	0
0.4	967	23 467	5354	929
0.8	3700	54 556	11 461	21 259
1.2	6351	127 194	20 592	411 408
1.6	9777	203 836	28 690	607 222
2.0	11 826	223 155	34 831	734 507
2.4	12 272	281 183	41 784	894 471
4.0	18 567	411 329	60 279	1 226 669
4.8	26 464	597 527	91 865	1 826 669
Fitted line	$Y = 5266X - 72$	$Y = 120256X - 16239$	$Y = 17870X - 1391$	$Y = 378000X - 86495$
$R^2$	0.9782	0.9803	0.9765	0.9616

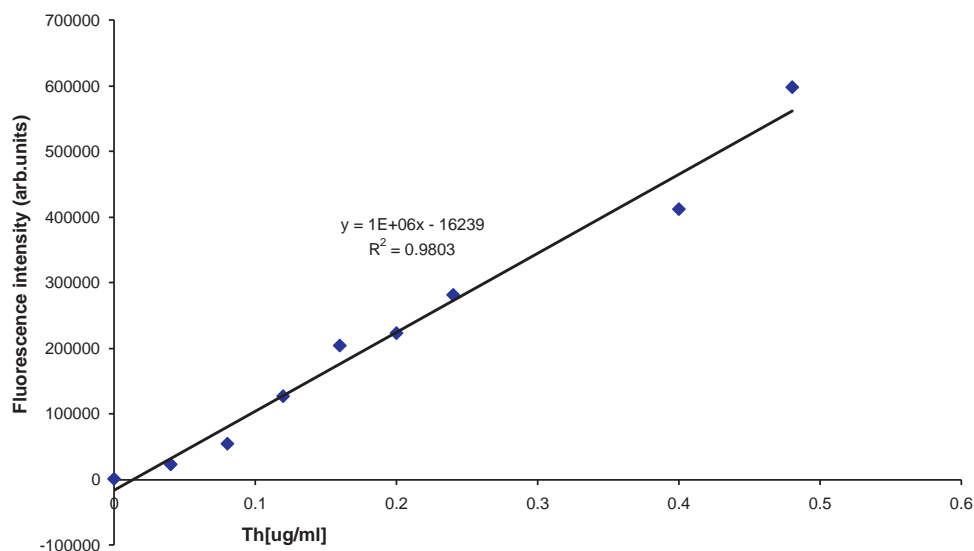


Fig. 3. Calibration of fluorimeter using Th-3-hydroxy flavone complex.

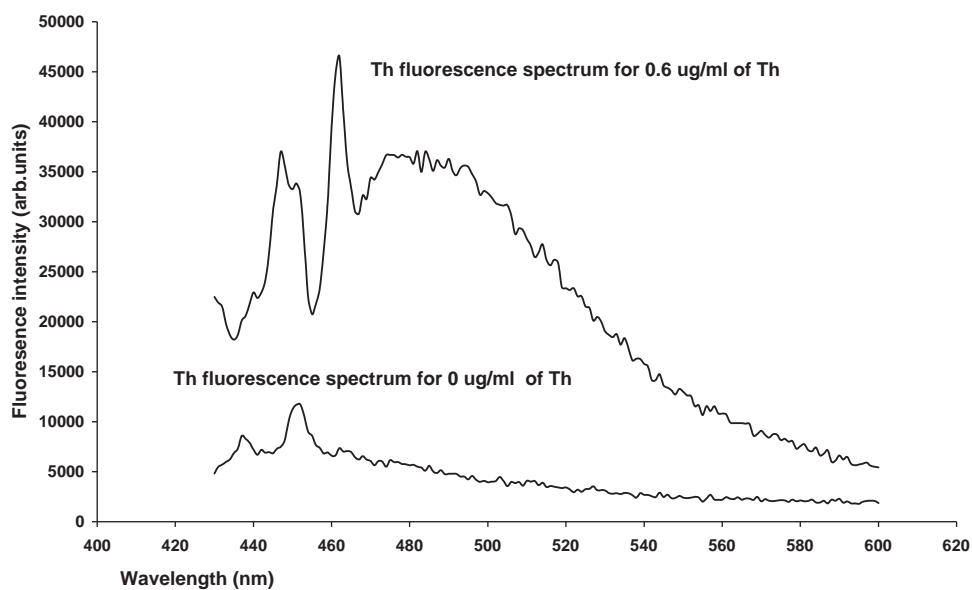


Fig. 4. Thorium-3-hydroxy flavone emission spectrum.

Table 3

Comparison of percent recovery of natural thorium both by spectrophotometric and fluorimetric techniques.

Method	Urine sample code	Th added ( $\mu\text{g}$ )	Th recovered ( $\mu\text{g}$ )	Bias	Recovery (%)	
Anion exchange method	Sam-01	4.0	3.2	−0.20	80.0	Average recovery: $(83.3 \pm 4.1)\%$ RSD: 4.9% Analysis time: 4 days MDA: 5.8 mBq/l $B_r$ : −0.17 $S_B$ : 0.04
	Sam-02	4.0	3.4	−0.15	85.0	
	Sam-03	4.0	3.1	−0.23	77.5	
	Sam-04	4.0	3.5	−0.13	87.5	
	Sam-05	4.0	3.3	−0.18	82.5	
	Sam-06	4.0	3.5	−0.13	87.5	
Solvent extraction method	Sam-07	4.0	3.4	−0.15	85.0	Average recovery: $(83.8 \pm 4.7)\%$ RSD: 5.6% Analysis time: 4 days MDA: 0.32 mBq/l $B_r$ : −0.17 $S_B$ : 0.05
	Sam-08	4.0	3.6	−0.10	90.0	
	Sam-09	4.0	3.3	−0.18	82.5	
	Sam-10	4.0	3.1	−0.23	77.5	
	Sam-11	4.0	3.5	−0.13	87.5	
	Sam-12	4.0	3.2	−0.20	80.0	

$B_r$ , mean bias;  $S_B$ , standard deviation of mean bias.

exchange method is 83.3% with a standard deviation 4.1%. Analysis time is four days with MDA 0.72  $\mu\text{g/l}$  (5.8 mBq/l). Average recovery obtained by the present developed fluorimetric method is 83.8% with a standard deviation 4.7%. Analysis time is four days with MDA 0.054  $\mu\text{g/l}$  (0.32 mBq/l) as shown in Table 3. In  $\text{Th}_{\text{nat}}$ , Th-232 is in secular equilibrium with Th-228. Hence, MDA is given as sum of activity due to both the isotopes.

### 3.5. Evaluation of bias

As per ANSI N13.30 standard for performance criteria for radio bioassay [13], the mean bias ( $B_r$ ) and standard deviation of mean bias ( $S_B$ ) are calculated. The accepted criteria for  $B_r$  is  $-0.25 \leq B_r \leq +0.5$  and accepted criteria for  $S_B \leq 0.4$ . Values of the mean bias and standard deviation of the mean bias are given in Table 3 for both anion exchange and solvent extraction method. It may be seen that these values are well within ANSI criteria for both the methods.

### 3.6. Derivation of minimum detectable activity (MDA) of the method

Minimum detectable activity of the method is determined using the following formula:

$$\text{MDA} = \frac{3\sigma}{F} \times \frac{100}{R} \times \frac{TV}{V}$$

where  $\sigma$  is the standard deviation of average process blank value in terms of fluorescence signal,  $F$  is the fluorescence value due to 1.0  $\mu\text{g}$  of Th,  $R$  is chemical recovery,  $TV$  is the volume of the urine sample received and  $V$  is the volume of the urine sample taken for analysis. In this developed method, average process blank value for 10 samples is  $7500 \pm 1050$  ( $1\sigma$ ). Average fluorescence intensity obtained for 10 replicate measurements for 1.0  $\mu\text{g}$  of Th is 93 034 and percent chemical recovery ( $R$ ) for Th is 83.8%. The MDA thus obtained for natural thorium in the present method computed by using the above formula is 0.04  $\mu\text{g/l}$  (0.32 mBq/l). In  $\text{Th}_{\text{nat}}$ , Th-232 is in secular equilibrium with Th-228. Therefore, both the isotopes will be present in equal amount of radioactivity from the estimated chemical amount of  $\text{Th}_{\text{nat}}$  in urine using fluorimeter.

## 4. Conclusion

The reliability of the indigenously developed solvent extraction method for monitoring  $\text{Th}_{\text{nat}}$  in urine matrix is compared with standard anion exchange method and the results are in good agreement. The MDA of  $\text{Th}_{\text{nat}}$  obtained for the developed method is 0.32 mBq/l and the analysis time is 4 days. The method developed as described above can very well be employed in *in vitro* monitoring of low level of thorium in urine matrix. This method is dedicated for natural thorium estimation in urine matrix. In  $\text{Th}_{\text{nat}}$ , Th-232 is in secular equilibrium with Th-228. Therefore, both the isotopes will be present in equal amount of radioactivity from the excreted chemical amount of  $\text{Th}_{\text{nat}}$  in urine. From the estimated chemical amount of  $\text{Th}_{\text{nat}}$  in urine using fluorimeter, activity due to Th-232 is calculated based on its specific activity and the same amount of activity due to Th-228 would be present in the urine sample. For the dose assessment, activities due to Th-232 and Th-228 isotopes will be taken into account as per excreted chemical amount of natural thorium in urine. However, to measure both the isotopes precisely, alpha spectrometry is essential.

## Acknowledgement

Authors wish to express their sincere thanks to Dr. B. Venkattraman, Associate Director Radiological Safety and Environmental Group for his constant support and guidance.

## References

- [1] ICRP, Individual Monitoring for Internal Exposure of Workers: Replacement of Publication 54, ICRP Publication 78, vol. 27 (3–4), Elsevier, Oxford, 1999, p. 33.
- [2] P. Roth, E. Werner, I. Wendler, P. Schramel, Appl. Radiat. Isotopes 47 (9/10) (1996) 1055.
- [3] H.S. Dang, D.D. Jaiswal, C.M. Sunta, S.D. Soman, Health Phys. 57 (3) (1989) 393.
- [4] C. Hurtgen, J. Radioanal. Nucl. Chem. 248 (2) (2001) 477.
- [5] S.G. Sarkar, S.V. Bandekar, P.M. Dhadke, J. Radioanal. Nucl. Chem. 243 (2000) 807.
- [6] D.K. Singh, H. Singh, C.K. Gupta, J.N. Mathur, J. Radioanal. Nucl. Chem. 250 (2001) 123.
- [7] R.K. Singh, P.M. Dhadke, J. Radioanal. Nucl. Chem. 254 (2002) 607.
- [8] R.G. Milkey, M.H. Fletcher, J. Am. Chem. Soc. 79 (1957) 5425.
- [9] G.G. Guilbault, Practical Fluorescence Theory Methods and Techniques, Marcel Dekker, New York, 1973.
- [10] R.S. Bottei, S.D. Alessio, Anal. Chim. Acta 37 (1967) 405.
- [11] N.A. Lange, Handbook of Chemistry, McGraw-Hill, Inc., 1956.
- [12] M.A. Gautier, Manual of analytical methods for radiobioassay, Los Alamos, in: LA-9763-M Manual, National Laboratory, New Mexico, 1983, R-210.
- [13] ANSI N13.30, Standard Performance Criteria for Radiobioassay, 1995.